Radiation doses of yttrium-90 citrate and yttrium-90 EDTMP as determined via analogous yttrium-86 complexes and positron emission tomography

Frank Rösch^{1,*}, Hans Herzog², Cornelius Plag², Bernd Neumaier¹, Ulrike Braun², Hans-Willhelm Müller-Gärtner², Gerhard Stöcklin¹

¹ Institut für Nuklearchemie, Forschungszentrum Jülich, Jülich, Germany ² Institut für Medizin, Forschungszentrum Jülich, Jülich, Germany

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Abstract. Yttrium-90 is used for palliative therapy for the treatment of skeletal metastases, but because it is a pure β - emitter, data on the pharmacokinetics and radiation doses to metastases and unaffected organs are lacking. To obtain such data, the present study employed yttrium-86 as a substitute for ⁹⁰Y, with detection by positron emission tomography (PET). The study compared the properties of two different ⁸⁶Y complexes - ⁸⁶Y-citrate and ⁸⁶Y-ethylene diamine tetramethylene phosphonate (EDTMP) - in ten patients with prostatic cancer who had developed multiple bone metastases (the ten patients being divided into two groups of five). Early dynamics were measured up to 1 h post injection (p.i.) over the liver region, followed by subsequent whole-body PET scans up to 3 days p.i. Absolute uptake data were determined for normal bone, bone metastases, liver and kidney. Radiation doses were calculated according to the MIRD recommendations. Based on the pharmacokinetic measurements of the distribution of the ⁸⁶Y complexes, it was possible to calculate radiation doses for the bone metastases and the red bone marrow delivered by complexes containing 90Y. In 1 cm³ of bone metastasis, doses of 26±11 mGy/MBq and 18±2 mGy/MBq were determined per MBq of injected 90Y-citrate and 90Y-EDTMP, respectively. The doses to the bone marrow were 2.5 ± 0.4 mGy/MBq for 90Y-citrate and 1.8±0.6 mGy/MBq for 90Y-EDTMP. 86Y and PET provide quantitative information applicable to the clinical use of ⁹⁰Y. This method may also be useful for the design of other 90Y radiopharmaceuticals and for planning radiotherapy dosages.

Key words: Yttrium-86 – Yttrium-90 – Positron emission tomography – Bone metastases – Quantitative in vivo radiation dosimetry

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Introduction

Approximately 80% of patients with prostatic carcinoma develop metastatic bone disease and nearly half of them experience bone pain [1]. Among the modalities for management of bone pain, endoradionuclide therapy (ERT) is a well-established approach. It uses compounds having an affinity for bone that are labelled with beta-emitting radionuclides. For some time, phosphorus-32, strontium-89 and yttrium-90 have been available as ³²P-phosphate, ⁸⁹Sr-chloride and ⁹⁰Y-citrate [2–4]. Chelates of phosphonates of samarium-153 and rhenium-186, namely, ¹⁵³Sm-EDTMP (ethylene diamine tetramethylene phosphonate) and ¹⁸⁶Re-HEDP (1,1-hydroxyethylidine diphosphonate) are reported to have therapeutically useful biodistributions.

The radiation doses to bone metastases as well as to normal bone and radiosensitive organs such as the bone marrow depend on the activity administered and on the pharmacokinetics of the radiopharmaceutical. The accurate determination of these radiation doses is a critical step in the development of a radiotherapeutical. Usually, measuring the uptake kinetics in individual organs and transferring these time-activity data to the MIRD calculation scheme provides the necessary dosage data. For humans such data are available qualitatively in the case of those β --emitting radionuclides that simultaneously emit a measurable percentage of low-energy photons. Samarium-153 and rhenium-186 are two examples; their decay generates photons of 103 and 137 keV in abundances of 28% and 9%, respectively. Consequently, some target to non-target accumulation ratios were deduced for ¹⁵³Sm-EDTMP by means of whole-body gamma camera images. Radiation doses were subsequently derived by Logan et al. [5], Eary et al. [6] and Bayouth et al. [7].

⁹⁰Y is a pure β --emitter without any accompanying γ -radiation. It is available in a convenient generator system for no-carrier-added syntheses and is efficiently incorporated into radiopharmaceuticals. Since 1990 more

Correspondence to: F. Rösch

^{*} Present address: Institut für Kernchemie, Johannes Gutenberg-Universität Mainz, D-55099 Mainz, Germany

than 50 papers have described ⁹⁰Y radiotherapy. There are even new initiatives to magnetically enhance ⁹⁰Y ERT [8].

To date, the calculations of the human doses of radiation from ⁹⁰Y radiopharmaceuticals have been indirect, based on one of several empirical approaches:

1. Extrapolating to man pharmacokinetic measurements made on animals ex vivo

2. Calculating from measurements of urinary excretion and blood clearance, based on the assumption that unexcreted radioactivity resides exclusively in skeletal metastases [9, 10]

3. Measuring human uptake kinetics by means of bremsstrahlung registration [11–14]

4. Substituting γ -emitting isotopes such as ⁸⁷Y or ⁸⁸Y for ⁹⁰Y and measuring radioactivity by means of gamma scintigraphy [15]

5. Extrapolating from pharmacokinetic data on indium-111 radiopharmaceuticals for the dosimetric calculations of the chemically similar ⁹⁰Y radiopharmaceuticals [16–21].

Recently an alternative approach was proposed to use PET to quantitatively assess the pharmacokinetics of ⁹⁰Y radiopharmaceuticals in humans, using the positronemitting isotope 86 Y ($T_{1/2}$ =14.74 h, 32% β^+) [22]. The present paper describes the application of this technique for the determination of uptake kinetics and the individual radiation dose of two ⁹⁰Y radiopharmaceuticals used in the treatment of disseminated bone metastases, namely, 90Y-citrate and 90Y-EDTMP. Although 90Y-citrate has been available for three decades, data from humans on its biodistribution and radiation doses of individual organs are not available. 90Y-EDTMP is not yet routinely used. It has great potential usefulness, however, since phosphonate ligands such as EDTMP promote the selective uptake of trivalent metallic radionuclides in bone and bone metastases. ¹⁵³Sm-EDTMP, for instance, is currently under clinical trial in the United States. Sm(III) and Y(III) have very similar co-ordination chemistry and charge-to-radius-ratios that allow substitution for Ca(II) in the hydroxyapatite of bone. Recently, clinical studies were reported on the pharmacokinetics of ⁹⁰Y-EDTMP in rabbits as well as in humans [23].

Materials and methods

Patients. Ten men with prostatic carcinoma and scintigraphically proven bone metastases were studied. Their ages ranged from 54 to 84 years. The patients had normal haematological parameters. Bone scintigrams with technetium-99m methylene diphosphonate were available for all subjects. None had received either radiotherapy or chemotherapy in the preceding 4 weeks. All had normal kidney and liver function. Prior to the study each patient gave written informed consent. Five patients each were chosen for investigation with ⁸⁶Y-citrate and ⁸⁶Y-EDTMP.

Preparation of ⁸⁶Y compounds. ⁸⁶Y was produced at the Jülich compact cyclotron CV28 via the (p,n)-reaction on 96.3% enriched

⁸⁶SrCO₃ target material; chemical processing led to no-carrieradded and radiochemically pure ⁸⁶Y(III) [24, 25].

The ⁸⁶Y-citrate and ⁸⁶Y-EDTMP complexes were synthesised by adding one drop of ⁸⁶Y(III) stock solution (10^{-4} *M* HCi) to ligand solutions of the following composition: (a) for citrate 3 ml aqueous solution of 7.5 mg/ml sodium citrate, pH 7.4, overall sodium citrate concentration 0.087 *M*, overall ionic strength 0.17 *M*, and 2 ml saline was added to obtain isotonicity; (b) for EDTMP 2 ml aqueous solution of 21.8–36.5 mg/ml EDTMP adjusted with NaOH to pH 7.5, and 1 ml saline was added to obtain isotonicity. The mixed solutions were sterile filtered and assays of pyrogenicity were performed by standard methods. Radiochemical quality control was performed by paper electrophoresis (Tris acetate, pH 6, as electrolyte) for ⁸⁶Y-citrate and by paper chromatography on MN 261 paper strips using pyridine/ethanol/water (1:2:4) mixtures, pH 7.4, for ⁸⁶Y-EDTMP. Radiochemical yields were found to be >98% for both complexes.

Radiopharmaceutical safety. The safety of ⁸⁶Y-citrate and ⁸⁶Y-EDTMP was ensured by using a sterile pyrogen-free preparation of the radiopharmaceutical, by monitoring the vital signs of each patient before and up to 1 h post injection (p.i.), by obtaining preand post-injection haematological profiles (white cells, red cells, platelet counts) and by pre- and post-injection radiochemical analysis of the compounds (blood, urine).

Radiopharmacokinetic studies. Patients had an intravenous line placed in each arm. Prior to the emission scans total-body transmission scans were recorded. The radiopharmaceuticals were administered intravenously in a volume of 4–5 ml. The administered activities for each subject are listed in Table 3. Dynamic PET scans were started simultaneously with the injection of the radiopharmaceutical and continued for 40–90 min. The field of view covered the liver and/or individual metastases in vertebrae. Later, up to four whole-body PET scans were acquired, the first at about 3 h p.i. and the last up to 76 h p.i.

To determine clearance from the blood, samples of venous blood were withdrawn at 1, 3, 5, 10, 20, 40, 60, 120 and 240 min p.i. The quantity of ⁸⁶Y excreted in the urine was measured periodically up to 24 h p.i. to obtain the total-body clearance by urinary excretion. Radioactivity in 1-ml aliquots of blood and urine was measured by γ -spectrometry. The activity in the blood pool at each time point was calculated from the measured activity per unit volume in the blood samples and the estimated blood volume [26]. If blood clearance kinetics showed two different phases, the first (fast) and second (slow) phases were denoted with half-lives of $T\alpha_{1/2}$ and $T\beta_{1/2}$. Results for $T\alpha_{1/2}$ refer to the periods from 0 to 5 min and 0 to 4 min p.i. for ⁸⁶Y-citrate or ⁸⁶Y-EDTMP, respectively, while those for $T\beta_{1/2}$ refer to the periods from 60 to 240 min and 20 to 240 min p.i., respectively.

The product of the radioactivity per unit volume of urine multiplied by the volume of that specimen, summed over all specimens, indicated the cumulated clearance of ⁸⁶Y-citrate or ⁸⁶Y-EDTMP. The activity in the whole body, $A_{\rm WB}(t)$, was obtained as the difference between whole-body activity and cumulative urinary excretion, $A_{\rm U}(t)$. This is represented as $A_{\rm WB}(t)=A_{\rm O}-A_{\rm U}(t)$, where $A_{\rm O}$ is the injected activity.

Moreover, blood and urine samples were analysed radiochemically by standard methods. After centrifugation, aliquots of the samples were transferred to a Sephadex G-25 column (20×2 cm). Radioyttrium fractions bound to serum proteins were chromatographically separated from the low-weight radioyttrium complexes using TRIS acetate (0.075 *M*, pH 7) as eluent. Elution profiles were recorded by UV as well as by radioactivity detectors. Individual fractions were subsequently collected and the amount of the two fractions was determined in percent.

PET measurement and data analysis. PET measurements were done with the Scanditronix/GE scanner PC4096-15WB [27]. As whole-body transmission scans were recorded before the first PET scan, all emission data could be corrected for attenuation. The patient was very carefully positioned for each measurement by means of laser beams in order to match the position of the transmission scan. The whole-body scans consisted of a series of single scans, each lasting 4–6 min, extending from the neck to the femur. In order to limit the time of investigation, other parts were imaged only if the bone scintigrams showed metastases. Depending on the extent of the whole-body scan, the reconstruction of the emission data yielded up to 210 transverse images for each measurement. These images could be reconstructed as anterior, lateral, coronal or sagittal sections.

The images had the unit ⁸⁶Y activity concentration, i.e. kBq/cm³. These primary data were corrected for the β^+ -abundance of 32% of ⁸⁶Y. This accuracy of the activity determination in structures not confounded by the partial volume effect was validated by phantom measurements. The contribution of the few γ -lines of ⁸⁶Y seen within the energy window of the scanner was satisfyingly corrected. The activity concentration in kBq/cm³ in the metastases was directly obtained by analysing regions of interest (ROIs) in the transverse slices, using a 30% isocontour level in three adjacent slices, and averaged taking into account the ROI areas. Furthermore, ROIs were marked over normal bone, liver and kidneys. The activity concentration for the liver is given as the mean of at least two different ROIs and that for the kidneys as the mean of one ROI per left and right kidney.

Finally, all data were corrected for the decay of ⁸⁶Y. Thus, the corrected data served for calculations of the absolute uptake kinetics of the ⁸⁶Y radiopharmaceuticals. Radionuclide concentrations are expressed in absolute units of kBq/cm³ and, in some cases, in relative units of %ID/cm³ or %ID/organ.

Calculation of radiation dose. Calculation of the radiation dose to the metastases, the red marrow, the liver and the kidneys was performed as described in detail elsewhere [22], in accordance with MIRD pamphlets no. 11 and no. 5 [28, 29]. The decay-corrected pharmacokinetic data of the ⁸⁶Y radiopharmaceuticals were considered to be valid for the analogous ⁹⁰Y-labelled compounds. A dose constant of 0.54 g·Gy/MBq·h (=2 g·rad/µCi·h) for ⁹⁰Y and a specific density of bone of 1.5 g/cm³ were used. The radiation dose to the red marrow was considered to be caused by activity equally stored in cortical and trabecular structures of healthy bone, i.e. the corresponding individual MIRD radiation dose Sfactors delivered from ⁹⁰Y stored in cortical and trabecular bone were applied.

Using the experimental data on excreted urine, the residence time for activity in the bladder content was determined as suggested in example 7 of the MIRD Primer [30] assuming voiding intervals of 2 h and 4 h, respectively.

In order to calculate the cumulated activities for both normal bone and metastases, the activity was considered not to be released after the uptake processes, which are completed at about 40 h p.i. for ⁸⁶Y-citrate and at about 2 h p.i. for ⁸⁶Y-EDTMP. This assumption is supported by our measurements over 3 days. This means that the short early periods of uptake can be negelected for the calculation of the cumulated activities. The activities averaged at 48 and 72 h for both ⁸⁶Y-citrate and ⁸⁶Y-EDTMP, respectively, were regarded as final levels of $A_{\text{metastasis}}$. With a biological decay constant of zero, the effective half-life equals the physical

half-life of 90 Y ($T_{\frac{1}{2}}$ =64 h), so that the cumulative activity is $\tilde{A}=A_0.64$ h·ln2.

The individual radiation doses are reported as mean ± 1 standard deviation. Due to the uncertainty in the parameters comprising the absorbed dose calculations, not more than two significant figures were used in reporting the absorbed dose results.

Results

PET imaging

Figure 1 shows typical transverse images displaying normal spine and bone metastases in a patient injected with ⁸⁶Y-EDTMP. Figures 2 and 3 compare typical wholebody views (recorded at 22 h p.i.) of ⁸⁶Y-citrate and ⁸⁶Y-EDTMP in anterior and midline sagittal projections. Similar results were obtained in the other patients of each group. Both radiopharmaceuticals show the normal skeleton and disseminated bone metastases as hot spots. However, the images produced by the two radiopharmaceuticals differ significantly: there is less soft tissue background in the ⁸⁶Y-EDTMP images and only the ⁸⁶Ycitrate images show the liver.

Pharmacokinetics

Early kinetics. Figure 4 shows the early phase of ⁸⁶Y-citrate uptake (0-60 min p.i.) in patient 4. Initially, a 10cm-broad body section over the liver was monitored that included some parts of the kidneys as well as a metastasis in the os ilium. Thus, these images served for measurements of the individual uptake kinetics of bone metastases, normal bone and liver and kidneys. 86Y accumulated continuously in bone metastases and also in normal bone, though at a much lower level. ⁸⁶Y-citrate concentrated in the liver rapidly, reaching a maximum of about 20%ID at 2-3 min p.i. The isotope was then released with a mean half-life of $T_{\frac{1}{2}}=90\pm20$ min. The kinetics of uptake by and release from the kidneys was similar to that for the liver. Maximum accumulation in the kidneys occurred somewhat later than in the liver, at about 3-5 min p.i. Within the first hour p.i., the release from this organ occurred with a $T_{1/2}$ =80±30 min.

Figure 5 shows the early uptake kinetics of ⁸⁶Y-EDT-MP in patient 6. Uptake by bone metastases was faster than that of ⁸⁶Y-citrate. Absolute values of ⁸⁶Y activity concentration varied significantly for individual bone metastases. In the kidneys, ⁸⁶Y uptake reached a maximum at about 2–5 min p.i., followed by a biphasic release consisting of a fast component having a $T\alpha_{1/2}=15\pm5$ min that lasted for up to 15 min p.i. and a slower component having a $T\beta_{1/2}=50\pm10$ min evident between 15 and 90 min p.i. Unlike following the injection of ⁸⁶Y-citrate, the liver showed a much lower accumulation of ⁸⁶Y-EDTMP.



Fig. 1a–d. Transverse images of 6.5-mm thickness recorded 4 h after injection of ⁸⁶Y-EDTMP in patient 10. Uptake is demonstrated in different bone metastases in the upper thorax (**a**), and in healthy spine (**b**), thoracic spine (**c**) and pelvis (**d**)



Fig. 2. Whole-body images of the distribution of ⁸⁶Y-citrate in patient 1 at 22 h p.i. *Left:* Anterior view with all coronal sections summed. *Right:* Sagittal section of 8-cm thickness in the median plane of the patient's body



Fig. 3. Whole-body images of the distribution of 86 Y-EDTMP in patient 10 at 22 h p.i. See legend to Fig. 2 for further explanation



Fig. 4. Decay-corrected initial time-activity curves of the early uptake kinetics of ⁸⁶Y-citrate in bone metastases, liver and kidneys (patient 4). *Bars* show deviations derived from ROI determinations for an individual metastasis, from two different ROIs for the liver, and from individual ROIs per left and right kidney

Overall kinetics. The overall uptake kinetics covers a period of up to 3 days p.i. Figures 6 and 7 show complete time-activity curves for all the individual bone metastases of patient 4, who received ⁸⁶Y-citrate, and of patient 6, who received ⁸⁶Y-EDTMP. The curves for ⁸⁶Y-EDT-MP are typical for all the patients of that group. After correction for decay, the activity concentration in bone metastases was essentially constant as early as ≈ 1.5 h



Fig. 5. Decay-corrected initial time-activity curves of the early uptake kinetics of ⁸⁶Y-EDTMP in bone metastases, liver and kidneys (patient 6). *Bars* show deviations derived from ROI determinations for two individual metastases, from two different ROIs for the liver, and from individual ROIs per left and right kidney



Fig. 6. Complete decay-corrected time-activity curves of various bone metastases, for ⁸⁶Y-citrate (patient 4)

p.i. In contrast, and also typical for the ⁸⁶Y-citrate group, activity concentrations in the metastases increased more slowly and reached final levels at approximately 2 days p.i. Average values of the uptake of ⁸⁶Y-citrate per cm³ of bone metastases as well as the (total) organ uptake for the skeleton and for the liver are summarised in Table 1. Importantly, the maximum activity concentrations in the ⁸⁶Y-citrate group were about 50% higher than those in the ⁸⁶Y-EDTMP group. For both radiopharmaceuticals, once the ⁸⁶Y activity concentration reached saturation it did not decline, indicating negligible clearance from the metastases.



Fig. 7. Complete decay-corrected time-activity curves of various bone metastases, for ⁸⁶Y-EDTMP (patient 6)

Table 1. Individual uptake of the two ⁸⁶Y complexes

Radioyttrium complex ligand	Time p.i. (h)	Uptake in various organs (%ID)				
		Bone metastases (per cm ³)	Skeleton (total)	Liver (total)		
Citrate	4	0.050±0.018	25.8±4.5	13.6±2.0		
	20	0.071±0.028	34.2±5.7	13.9±2.0		
	48	0.085±0.037	36.2±5.9	11.2±1.8		
	72	0.102 ± 0.013	44.0 ± 1.4	10.0 ± 2.2		
EDTMP	4	0.051±0.004	23.5±8.0	а		
	20	0.050 ± 0.004	24.2±7.7	а		
	48	0.054 ± 0.008	23.5±7.8	а		
	72	0.053±0.003	23.8±2.7	а		

^a At the background level

Although the kinetics of uptake were similar within each group, the maximum ⁸⁶Y activity concentrations, expressed as %ID/cm³, varied not only between patients but also between metastases in the same patient (cf. Fig. 7 for example). Average values of the uptake of both the ⁸⁶Y-complexes are summarised in Table 1.

The different uptake kinetics of 86 Y-citrate and 86 Y-EDTMP by bone metastases in turn determine the ratio of 86 Y activity concentration ratios between bone metastases and normal bone. In a given patient these ratios varied significantly, from 3:1 to 15:1. Similar ratios of 8 ± 2 were obtained at about 1 h p.i. for 86 Y-EDTMP and at about 2 days p.i. for 86 Y-citrate. There is evidence that the 86 Y uptake kinetics in normal bone is somewhat faster than in bone metastases. The healthy bone seems to be more rapidly covered by 86 Y to a final level. The bone metastases may increasingly incorporate 86 Y due to the weaker apatite matrix.

Table 2. Blood clearance and urmary whole-body retention da	Table	2. Blood c	learance and	urinary	whole-body	retention	data
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⁸⁶ Y complex	Patient no.	Ligand concentration	Blood clearance					Whole-body retention
			$T\alpha_{\frac{1}{2}}$ (min)	%ID 10 min p.i.	Time for 50% clearance (min)	%ID 2 h p.i.	$T\beta_{\frac{1}{2}}$ (min)	4 h p.i. (cumulated)
Citrate ^a	3	Isotonic	1.9	16.7	1.5	1.1	90	91
	4	Isotonic	6.3	51.6	11.8	5.9	105	84
	5	Isotonic	4.3	33.3	4.5	2.1	99	98
	Mean		4.2±1.8	34±14	5.9±3.1	3.0±2.1	98±6	91±6
EDTMP	6	43.5 mg	2.1	11.3	1.1	0.7	65	93
	7	75.0 mg	1.3	6.5	1.0	0.3	61	(18)
	8	45.6 mg	2.2	25.4	1.5	1.3	59	65
	9	48.9 mg	6.5	43.3	7.6	4.7	150	72
	10	46.0 mg	2.1	19.0	1.1	3.5	70	64
	Mean	-0	2.8±0.9	21±6	2.5±1.3	2.1±0.9	81±38	73±13 ^b

^a Blood pool and urinary excretion was not measured in patients 1 and 2

^b Calculated without patient 7 because of the higher EDTMP concentration injected



Fig. 8. Average blood clearance kinetics of ⁸⁶Y-citrate and ⁸⁶Y-EDTMP. *Bars* show standard deviations (n=3 for ⁸⁶Y-citrate, n=5 for ⁸⁶Y-EDTMP)

For ⁸⁶Y-citrate, the ⁸⁶Y activity concentration in the liver peaked at 2–3 min p.i. and then decreased continuously over the whole period of measurement. At 20 h p.i. and 72 h p.i. this organ contained about 14% and 10%, respectively, of the total administered activity (Table 1).

Blood clearance and whole-body retention

Table 2 summarises individual blood clearance and whole-body retention data. Average blood clearance and average whole-body retention kinetics are summarised in Figs. 8 and 9. The clearance of ⁸⁶Y from blood was biphasic, consisting of a rapid phase lasting about 10 min p.i., followed by clearance at a slower rate (Table 2). An average of 34%±14% of the injected ⁸⁶Y activity re-



Fig. 9. Average whole-body retention kinetics of 86 Y-citrate and 86 Y-EDTMP. *Bars* show standard deviations (*n*=3 for 86 Y-citrate, *n*=4 for 86 Y-EDTMP)

mained in the blood at 10 min after the injection of ⁸⁶Ycitrate, compared with only 21%±6% of the ⁸⁶Y-EDTMP activity. The half-times of the rapid clearance phases were $T\alpha_{\frac{1}{2}}$ =4.2±1.8 min for ⁸⁶Y-citrate and 2.8±0.9 min for ⁸⁶Y-EDTMP. Table 2 also includes the time when 50% of the radioactivity was removed from the blood. At the end of 1 and 2 h, blood radioactivity had dropped to 11%±6% and 3.0%±2.1% for ⁸⁶Y-citrate and to 7.4%±3.9% and 2.1%±0.9% for ⁸⁶Y-EDTMP, the values of $T\beta_{\frac{1}{2}}$ for this slow phase being 98±6 min and 64±4 min, respectively.

The whole-body retention of 86 Y-citrate by individual patients was very similar, $91\% \pm 6\%$ at 4 h p.i. For 86 Y-EDTMP the whole-body retention was $73\% \pm 13\%$ at 4 h p.i. with one exception. Interestingly, patient 7, who received the highest EDTMP dose of 75 mg, excreted 82%

Table 3. Individual radiation doses calcu-
lated for the injection of 1 MBq of the two
⁹⁰ Y complexes

Radioyttrium complex lígand	Patient no.	⁸⁶ Y activity injected (MBq)	Radiation dose (per MBq 90Y injected)			
			Bone metastases ^{a, c} (mGy/MBq)	Red marrow ^{b, d} (mGy/MBq)	Liver ^d (mGy/MBq)	
Citrate	1	225	22±8	2.0	1.8	
	2	130	25±7	3.0	1.6	
	3	295	35±1	2.5		
	4	265	36±3	2.9	2.1	
	5	180	10±1	2.3		
	Mean		26±11	2.5±0.4	1.8±0.3	
EDTMP	6	245	20±5	1.5		
	7	220	16±2	1.9		
	8	150	17±2	1.5		
	9	150	19±6	1.2		
	10	195	17±3	2.7		
	Mean		18 ± 2	1.8±0.6		

^a Data could not be compared by t test. A rank sum test showed no significant difference

^b A t test revealed a significant difference (P<0.05) between ⁸⁶Y-citrate and ⁸⁶Y-EDTMP

^c Radiation dose per one cm³ volume

^dRadiation dose per organ

ID by 4 h p.i., i.e. the whole-body retention was only 18%.

Biochemical analysis

Radiochemical analysis of blood samples showed that the stability of the two ⁸⁶Y radiopharmaceuticals differed in vivo. Gel chromatography revealed that 5 min after the injection of ⁸⁶Y-citrate, >60% of the ⁸⁶Y in the blood was bound to serum proteins. In contrast, the ⁸⁶Y-EDTMP complex was much more stable; at 5 min p.i., ⁸⁶Y bound to serum proteins accounted for only 7% of the blood ⁸⁶Y activity.

Radiation doses of ⁹⁰Y radiopharmaceuticals

Using the uptake kinetics measured with ⁸⁶Y, the radiation dose was calculated for the ⁹⁰Y analogues, cf. [22]. The individual patient data for the bone metastases, the liver and the red marrow are summarised in Table 3, which also includes mean values and standard deviations of the radiation doses for each radiopharmaceutical.

The average dose to 1 cm³ of bony metastases, per MBq ⁹⁰Y injected, would be greater for ⁹⁰Y-citrate (26±11 mGy/MBq) than for ⁹⁰Y-EDTMP (18±2 mGy/MBq) (P>0.05). ⁹⁰Y-citrate also would deliver a higher dose to red marrow (2.5±0.4 mGy/MBq) than ⁹⁰Y-EDTMP (1.8±0.6 mGy/MBq) (P<0.05) (Table 3). Due to the negligible uptake of ⁹⁰Y-EDTMP by liver, only the uptake of ⁹⁰Y-citrate is relevant, this would be 1.8±0.3 mGy/MBq.

Radiation doses to the kidneys were available for those patients in whom uptake kinetics of the two ⁸⁶Y-complexes for this organ were measured within the early

period of accumulation. For ⁸⁶Y-citrate, this was the case for patients 3, 4 and 5. According to the varying blood clearance kinetics (cf. Table 2), the residence times for these patients were different too. The calculated data were 4.11 min, 1.74 min and 0.62 min, respectively. This resulted in radiation doses of 0.12, 0.05 and 0.02 mGy/MBq, respectively, per MBq ⁹⁰Y-citrate injected. In the case of ⁸⁶Y-EDTMP one patient (no. 9) was analysed. The residence time determined was 1.37 min and the corresponding radiation dose amounted to 0.04 mGy/MBq for ⁹⁰Y-EDTMP.

Radiation doses to the bladder wall were calculated for patients 3–5 (⁸⁶Y-citrate) and patients 6–10 (⁸⁶Y-EDTMP). As these data were derived from the mean values of whole-body retention, cf. Table 2 and Fig. 9, average radiation doses were obtained for this organ in the case of both of the ⁹⁰Y complexes. Assuming voiding intervals of 2 h and 4 h, the radiation doses were 1.1 and 2.2 mGy per MBq of ⁹⁰Y-citrate injected, respectively, and 0.9 and 2.1 mGy per MBq of ⁹⁰Y-EDTMP injected.

Discussion

As expected, both the ⁸⁶Y radiopharmaceuticals localised primarily in the skeleton. Metastases accumulated ⁸⁶Y-EDTMP rapidly, attaining maximum concentrations by 1.5 h p.i. ⁸⁶Y-citrate accumulated much more slowly, requiring about 2 days to reach a maximum, but the level attained was higher than that achieved by ⁸⁶Y-EDTMP. Once deposited, the ⁸⁶Y of both radiopharmaceuticals remained in bone metastases or normal bone.

The individual bone metastases to normal bone accumulation ratios for ⁸⁶Y-citrate and ⁸⁶Y-EDTMP were similar, ranging between 3:1 and 15:1. However, final average ratios of 8 ± 2 for ⁸⁶Y-EDTMP were already reached at about 1 h p.i. and remained constant over the entire period of measurement. For ⁸⁶Y-citrate, on the other hand, the average ratio at about 1 h p.i. amounted to 6.5 ± 1.5 and increased to 8 ± 2 at 2 days p.i.

Data on the accumulation of ⁸⁶Y permitted calculation of the radiation doses that would be delivered by ⁹⁰Y radiopharmaceuticals. Such calculations indicate that ⁹⁰Y-citrate will deliver a slightly higher dose to 1 cm³ of metastases than ⁹⁰Y-EDTMP (26 ± 11 vs 18 ± 2 mGy/MBq ⁹⁰Y administered, respectively). The ROI technique of evaluating the activity uptake into a single metastasis is influenced by the partial volume effect. Therefore, it is expected that the activity in some parts of the metastasis as well as the related radiation dose will be higher than the data derived from the ROI mean.

The corresponding doses to total bone marrow per MBq administered 90 Y, as derived from the accumulation of 90 Y in healthy bone, will be 2.5±0.4 and 1.8±0.6 mGy/MBq. Owing to preferential deposition of the radiopharmaceuticals in bone metastases, the local dose to bone marrow in the vicinity of a metastasis could be up to 15-fold higher. This, in turn, will increase the average radiation dose to the bone marrow.

The stabilities of the Y-citrate and Y-EDTMP complexes significantly influence their distributions in the body, their rates of accumulation in bony metastases and, finally, the radiation doses delivered to such metastases. Y-EDTMP is more stable than Y-citrate, the ig β of the complexes being ≈ 20 and 7.9, respectively [31–33]. Owing to the greater stability of the Y-EDTMP complex, the binding of yttrium to blood proteins and uptake by the liver are negligible, and a greater fraction of the injected dose is immediately available for uptake by bone and bony metastases. The remainder of the ^{86/90}Y is rapidly excreted via the kidneys.

Some parts of the Y³⁺ released by the dissociation of the citrate complex bind to serum and are taken up by the liver. The sequestration and subsequent slow release of yttrium from these depots accounts for the slow accumulation of the radionuclide in bone and for the greater whole-body radiation dose. On the other hand, it prevents a significant portion of ^{86/90}Y from being excreted rapidly. This is reflected in the whole-body retention of the two ^{86/90}Y complexes. In turn, this intermediate storage allows for redistribution of ^{86/90}Y, providing a steady transfer of the radionuclide from the liver to bone metastases. Consequently, this process is an explanation for the higher activity concentration of ⁸⁶Y in the bone metastases obtained in the case of ⁸⁶Y-citrate. The results of the studies of ⁸⁶Y binding to serum proteins in the present study, as well as studies of the biodistribution of complexes of ⁸⁷Y(III) with citrate and EDTMP in rats [34], support this interpretation. The replacement of Ca(II) in the hydroxyapatite lattice by Y(III) in the form of Y³⁺ cation is an important mechanism for the deposition of yttrium in bone. For 86/90Y-EDTMP the binding of the phosphonate moiety to the hydroxyapatite matrix is also important [35]. In both of these processes, the

stability of the Y-EDTMP complex would tend to limit the amount of yttrium incorporated and thus could contribute to the lower radiation dose delivered by this radiopharmaceutical.

Conclusion

The clinical application of the β -emitter ⁹⁰Y in endoradiotherapy requires quantitative data on both the biodistribution and the radiation dose of 90Y radiopharmaceuticals. Studies employing β^+ -emitting ⁸⁶Y as a substitute for β --emitting ⁹⁰Y and direct in vivo measurement by PET compared the pharmacokinetics and radiation dosages of 86/90Y-citrate and 86/90Y-EDTMP complexes, each in groups of five patients with bone metastases from carcinoma of the prostate. Owing to the stability of the ^{86/90}Y-EDTMP complex, sequestration of ^{86/90}Y in blood and liver was negligible, and concentrations of ^{86/90}Y in bone metastases reached a maximum by 1.5 h. Because a substantial fraction of the 86/90Y delivered by the citrate complex is taken up by and then slowly released from the liver, concentrations in metastases required about 2 days to reach a maximum in the case of 86/90Ycitrate but bone metastasis levels were higher than those achieved with 86/90Y-EDTMP.

The results of the present study suggest that this technique could be superior to the other methods used to estimate in vivo radiation doses of ⁹⁰Y radiopharmaceuticals. Moreover, the study demonstrates the feasibility of using this technique to compare different ⁹⁰Y radiopharmaceuticals in vivo.

Finally, a preliminary ⁸⁶Y PET trial in an individual patient can serve as a precise basis for calculating the dose of ⁹⁰Y to be used subsequently in radiotherapy. In determining the activity of ⁹⁰Y-citrate and ⁹⁰Y-EDTMP that can be safely administered to a patient, the radiation dose to both bone metastases and the red marrow should be taken into consideration.

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